

Abnormality Detection in Ovarian Ultrasound Images using Active Contours

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ABSTRACT

Analyzing ovarian size parameters and its shape is the first hand information required in Ovary diagnosis. Automated segmentation of Ovary and measuring its size and analyzing its shape can help doctors in mass screening programme in faster diagnosis. In this paper, we propose a new improved algorithm (anisotropic diffusion filter, CLAHE enhancement, and active contour segmentation) to extract the Ovary from its homogeneous background followed by geometrical and shape feature extraction of the Ovary for automated computer-assisted analysis. The proposed algorithm is tested on 75 Transvaginal ultrasound images of ovaries. The experimental results are validated against the manual measurements done by the expert and the results obtained by our algorithm are in good agreement with experts inputs. The proposed algorithm could achieve an average Error Percentage EM_1 of 4% for Major-Axis length and average Error Percentage EM_2 of 5.43% for Minor-Axis length and classification rate of 91.66%.

Keywords: Image segmentation, Ovary, anisotropic diffusion filter, active contour, Geometric features, Ovary classification

1. INTRODUCTION

Ultrasound imaging has become an integral diagnostic tool to evaluate the gynecological health of a woman [1]. In this paper, we propose a new modified algorithm for computer-assisted analysis of the ultrasound images of women's ovaries. Ovary is scanned to diagnose abnormalities like infertility, irregular menstrual cycles, PCOS, cysts, tumors etc. Analyzing the ovarian size parameters and its shape are one of the first steps in diagnosis of the Ovary [1-3]. The normal range of Ovary size parameters is listed in Table I which was obtained in literature and supported by our expert Dr. Sriprada Vinekar, Gynecologist and Sonographer. Lists the parameters considered to evaluate the shape of the Ovary. Values listed in Table 1 are used to evaluate whether Ovary is normal or abnormal.

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Table 1: Ovary Condition

Normal Ovary	Min	Max
Major-axis length M1	3cm	5cm
Minor-axis length M2	1.5cm	3cm
Thickness	0.5cm	1.5cm
Area	3.54cm ²	11.8cm ²
Perimeter	7.45cm	12.95cm
Form Factor F	0.8	0.88
Compactness Q	14.2	15.6
Eccentricity E	>0.5	
Shape	Oval (Almond)	

2. BACKGROUND WORK

Automated measurement of ovarian parameters requires segmentation of the Ovary from the ovarian ultrasound images. The ultrasound images are of low contrast and poor quality due to the presence of speckle noise. Moreover the Region of Interest (Ovary) is highly homogeneous with its background. Hence, edge preserving speckle filters are required to improve the image quality and possibly the diagnostic potential of the image. In [1] Bilateral filter proposed by Tomasi C, Manduchi R[4] was used for speckle removal, Contrast Stretching and Power Law Transformation were used for enhancement, adaptive Thresholding and Global Thresholding for segmentation to extract Ovary. The average Error Percentage for Major-Axis length (E_{M1}) obtained was 11.9% and the average Error Percentage for Minor-Axis length (E_{M2}) obtained was 17.4%. In [2] anisotropic diffusion filter proposed by Perona and Malik [5, 6] was used for speckle reduction, CLAHE [7] followed by adaptive binary thresholding to extract Ovary. The algorithm could achieve an average Error Percentage of 5.27% for Major-Axis length (E_{M1}) and average Error Percentage of 6.1% for Minor-Axis length (E_{M2}). In [8, 9] active contours was used for detection of follicles. The average detection rate was 96.66%, false rejection rate (FRR) 3.33 %, and false acceptance rate (FAR) 15.04%. Active contours detect edges even in high noisy region and hence in this paper we propose to use active contours for segmentation.

The paper is organized as follows: Section III provides the description of the algorithm where speckle reduction filter, contrast enhancement methods, segmentation techniques and feature extraction are explained in detail. Section IV, the experimental results are discussed along with manual measurements done by the expert and section V contains conclusions and future scope of work.

3. MEASUREMENT OF OVARIAN FEATURES AND CLASSIFICATION

The input is a Transvaginal ultrasound image of Ovary acquired from GE-LOGIQ Book XP ultrasound machine which is a high performance multipurpose color hand-carried imaging system. The images are obtained in JPEG-LS (24-bit, 532x434x3) format. The sample ultrasound image is shown in Figure.1 consists of the Ovary region and Ovarian follicles inside the Ovary.

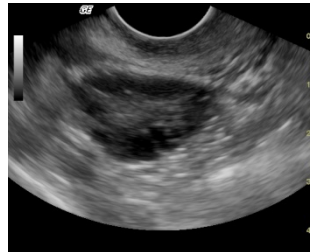


Figure. 1 Ultrasound Ovary image

The input image has identical RGB planes hence, only R-plane (8-bit, 532x434) is considered as the input image. The position of the Region of Interest, Ovary in all images is in the center of the fan area, so a sub-image I_{ROI} containing only the Ovary (ROI) of size [310 420] is extracted.

3.1 Preprocessing

The proposed Ovary feature extraction system is shown in Figure.2. Measurement of the size and shape features of the Ovary requires segmentation of the Ovary from the ultrasound image. Since ultrasound images are invariably noisy due to speckle noise, we use speckle reducing anisotropic diffusion filter for denoising. The edge magnitude parameter k was chosen as 0.9 and number of iterations fixed at 5. The filtered output I_{f2} has very good mean preservation, variance reduction, and edge localization.

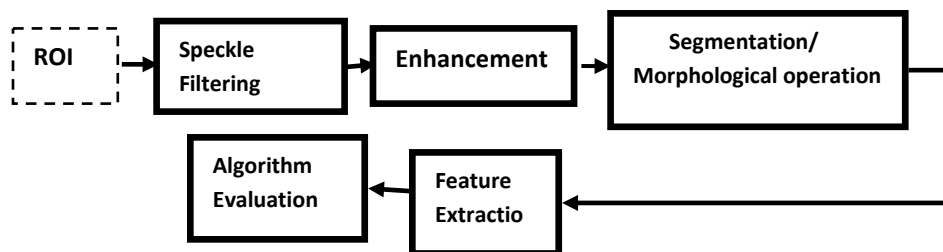


Figure.2 Ovary Feature Extraction System

To create a good contrast between the Ovary and the background which is essential for segmentation, Contrast Limited Adaptive Histogram Equalization (CLAHE) is applied. This algorithm operates on small data regions (tiles), instead of an entire image. Each tile's contrast is enhanced using Rayleigh's distribution. The neighboring tiles are combined using bilinear interpolation to eliminate artificially induced boundaries. In this algorithm we have chosen the tile value as [3 2], each tile's contrast is enhanced using Rayleigh's distribution. The clip-limit is

chosen as 0.1, range as original and alpha which is a distribution parameter is chosen as 6. The output of the enhancement stage $ICL(x,y)$ has a clear contrast between Ovary and background and is suitable for segmentation.

3.2 Processing

The processing stage has Segmentation, Morphological operation and feature extraction. The active contours without edges are used for segmentation. The resulting image after segmentation has many regions apart from Ovary. To begin with regions touching the image border are removed followed by region filling operation. We next perform morphological erosion (3 times with window size 3x3) followed by morphological dilation (3 times with window size 3x3). Many small phantom regions exist along with the Ovary region. A connected component labeling algorithm finds all connected components in the image and assigns a unique label to all points in the same component. Regions having smaller area than the threshold T are removed. This might result with more than one component. The Centroid ($C = C_x, C_y$) of the remaining components are found. In the current database, the Ovary is in the middle of the fan area and the mean centroid ($C = C_x, C_y$) of Ovary is experimentally found. Rule1 & Rule2 shown below are used to retain Ovary and eliminate the rest of the regions/components.

$$\text{Rule1: } \mu_{C_x} - k \sigma_{C_x} \leq C_x \leq \mu_{C_x} + k \sigma_{C_x}$$

$$\text{Rule2: } \mu_{C_y} - k \sigma_{C_y} \leq C_y \leq \mu_{C_y} + k \sigma_{C_y}$$

3.3 Feature Extraction

The outcome of the processing stage is a single component, Ovary. The sample input Ovary and the segmentation result is shown in Figure.3.

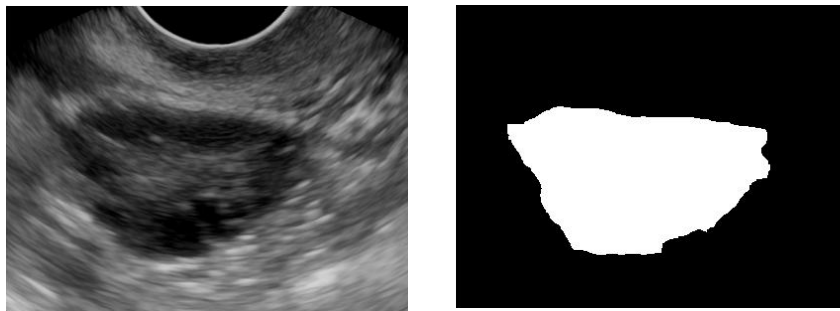


Figure.3 Segmentation result of Ovary

The geometrical and shape features of the segmented region are extracted which are grouped under F1 and F2 feature sets. The feature set F1: is the size based features and F2: is the shape based features [3].

F1: This feature set includes the Major-axis length $M1$, Minor-axis length $M2$, Area A and Perimeter P .

F2: This feature set includes Form Factor F, Compactness Q, and Eccentricity E.

3.4 Classification

In the training phase, the F1 and F2 feature sets of the ovaries are extracted. The mean and standard deviation of each of this parameter is calculated. The μ and σ of these parameters are listed in TABLE 2. Based on these values classification rules are formulated as shown in TABLE 3.

Table 2: mean and Standard deviations

Parameter	Mean, sigma
$\mu M_1, \sigma M_1$	3.459, 0.757
$\mu M_2, \sigma M_2$	1.987, 0.565
$\mu A, \sigma A$	5.494, 2.5936
$\mu P, \sigma P$	10.28, 2.82
$\mu F, \sigma F$	0.8451, 0.12
$\mu Q, \sigma Q$	15.15, 2.97
$\mu C_x, \sigma C_x$	191.32, 37.65
$\mu C_y, \sigma C_y$	110.77, 27.277

The classification rules of the $k\sigma$ -Classifier are formulated as in TABLE IV. A region is classified as a normal Ovary, if the following conditions Rule_i, $i=1,2,\dots,9$ are satisfied. The value of constant k is 2, α is 0.3 in the experiment. All the values are empirically determined.

Table 3: Rules

Rule1: $\mu C_x - k \sigma C_x \leq C_x \leq \mu C_x + k \sigma C_x$
Rule2: $\mu C_y - k \sigma C_y \leq C_y \leq \mu C_y + k \sigma C_y$
Rule3: $\mu M_1 - k/3 \sigma M_1 + \alpha \leq M_1 \leq \mu M_1 + k \sigma M_1 + \alpha$
Rule4: $\mu M_2 - k/3 \sigma M_2 + \alpha \leq M_2 \leq \mu M_2 + k \sigma M_2 + \alpha$
Rule5: $\mu A - k/2 \sigma A + \alpha \leq A \leq \mu A + k \sigma A + 2 \alpha$
Rule6: $\mu P - k/2 \sigma P + \alpha \leq P \leq \mu P + k \sigma P + \alpha$
Rule7: $\mu F - k \sigma F \leq F \leq \mu F + k \sigma F$
Rule8: $\mu Q - k \sigma Q \leq Q \leq \mu Q + k \sigma Q$
Rule9: $\mu E - k \sigma E \leq E \leq \mu E + k \sigma E$

In the testing phase, Feature set F1 and F2 are computed for the segmented region of Ovary and the classification rules Rule_i are applied to determine whether the Ovary is normal or not. Rule1 & Rule2 are applied to retain Ovary and eliminate other regions when multiple regions of same size are the segmentation outcome. Rule3 to Rule6 classifies the Ovary based on size and are the key rules for classifying. R8 to R9 are based on shape and support the Rule3-6.

3.5 Performance Measures

The accuracy of Ovary segmentation is measured using two parameters- the error percentage [1, 2] for Major-axis length (EM1) and Minor-axis length (EM2) which is used to evaluate and validate the algorithm. We calculate the percentage error between the expert measured value of M1 and M2 and algorithm evaluated value of M1 and M2. This parameter qualifies the algorithm as to how best or how near the algorithm is to the expert measured values. It is required that ideally the error percentage is "ZERO" but a ±12% deviation is acceptable as per the expert.

The error percentage [1, 2] for Major-axis length (EM1) and Minor-axis length (EM2) is calculated by using (1) and (2).

$$E_{M1} = \frac{|((\text{length})_{\text{obtained value}} - (\text{length})_{\text{Expert value}})|}{(\text{length})_{\text{Expert value}}} \times 100 \quad (1)$$

$$E_{M2} = \frac{|((\text{width})_{\text{obtained value}} - (\text{width})_{\text{Expert value}})|}{(\text{width})_{\text{Expert value}}} \times 100 \quad (2)$$

Ovary segmentation process, might result in more than one regions. So in order to select the Ovary we use Rule 1 & 2. But if there are more than one regions with in the centroid with allowable variance, algorithm might select a wrong region as Ovary. This might lead to misclassification. An Ovary is classified either as normal or abnormal. Consequently, there are four events; two classifications and two misclassifications.

3.5.1 Classification Rate

The classification rate or classification accuracy is estimated by the ratio of the total number of correctly classified Ovary images (sum of true positives and true negatives) by the number of input Ovary images.

$$\text{Classification Rate} = \frac{\text{\#of Correctly Classified Ovary Images}}{\text{\#of Input Ovary Images}} \quad (3)$$

3.5.2 False Acceptance Rate

False acceptance, also called a type II error, is defined as the percentage of abnormal Ovary images classified as normal.

3.5.3 False Rejection Rate

False Rejection Ratio, also called a type I error, is defined as the percentage of normal Ovary images misclassified as abnormal.

4. RESULTS AND DISCUSSIONS

The database consists of 75 Transvaginal ultrasound images of ovaries. The size parameter M1 and M2 are provided by the expert for all the 75 images. The classification output is validated against the expert inputs. In our experiment, 40 images are used in training phase and 35 (20 normal, 15 abnormal) in testing phase. The sample outputs where the edge of the segmented Ovary is superimposed on the input image for subjective validation by the expert are shown in Fig 4. The first column of images shows the expert measured values of Major-axis length M1 and Minor axis length M2 and the second column shows the output images. The F1 features and F2 features and classification output of the sample images are shown in Table VI. The average Error Percentage obtained is 4% for Major-Axis length (EM1) and 5.45% for Minor-Axis length (EM2) which is very much lesser than the BCAT method or ACAT method [3]. EM1 was 11.9% and EM2 was 17.4% using BCAT method whereas EM1 was 5.27% and EM2 was 6.1% using ACAT method. The average Error percentage using our new algorithm is very insignificant and proves to be better than ACAT and BCAT method.

The $K-\sigma$ classifier rules listed in TABLE IV is applied to the feature set F1, F2 to classify the Ovary as normal or abnormal. Out of 35 images used for testing, 20 images are normal and 15 images are abnormal. Classification results obtained by the algorithm is discussed below:

Among 20 normal Ovary images 18 are classified as normal and 2 are classified as abnormal. Among 15 abnormal images, 14 are classified as abnormal and 1 image is classified as normal. The classification rate is 91.66% and false acceptance rate (FAR) is 6.66% and false rejection rate (FRR) is 10% respectively.

The classification rate has significantly improved to 91.66% with our new algorithm compared to BCAT method in which classification rate was 76.67% and ACAT method in which classification rate was 85.8%. The TABLE 4 shows the classification results.

Table 4: Results

Methodology	Classification Rate	Type II Error (FAR)	Type I Error (FRR)
New Algorithm	91.66%	6.66%	10%
BCAT	76.67%	26.66%	20%
ACAT	85.8%	13.33%	15%

5. CONCLUSIONS

The classification rate depends on the choice of filter used, enhancement methods and segmentation method. The average Error Percentage is 4% for EM1 and 5.45% for EM2 which is significantly better than BCAT and ACAT method. From TABLE 5 it can be inferred that ACA has a good classification rate of 91.66%, FAR of 6.66% and FRR of 10%. The algorithm can be further improved using filters that can preserve edges better and segmentation methods that can extract the Ovary.

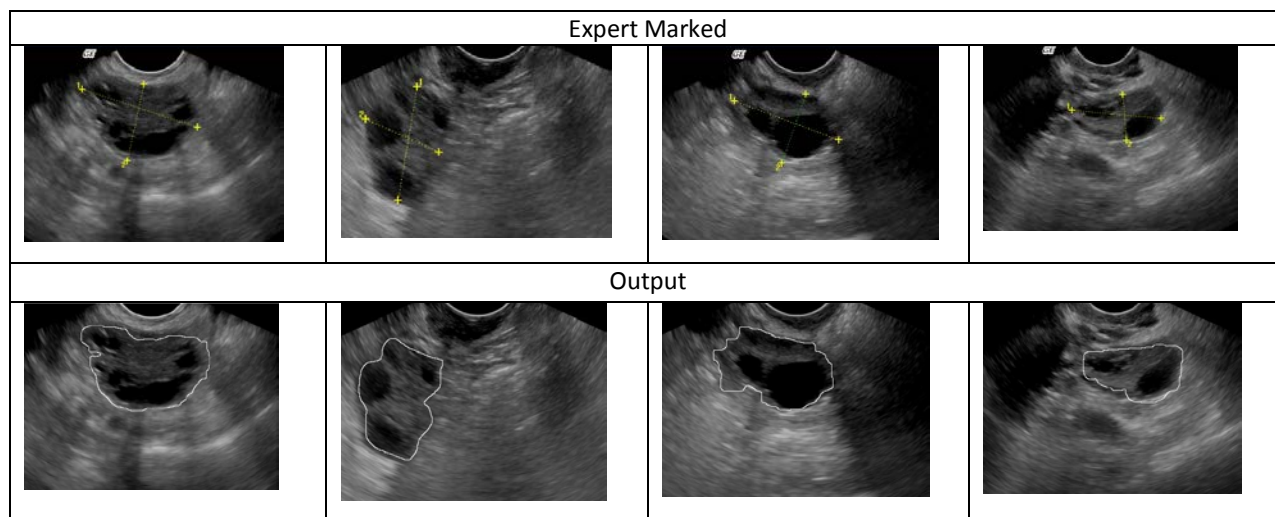


Figure 4. Segmentation results

Table 5: Classification Rates

#	M ₁ (Expert)	M ₂ (Expert)	M ₁ (Cm)	M ₂ (Cm)	A(Cm ²)	P(Cm)	F	Q	E	Classification
1	2.84	1.41	2.838	1.443	3.14	6.75	0.86	14.51	0.86	Normal
2	4.08	1.14	4.220	1.33	3.65	9.82	0.45	16.52	0.949	Normal
3	3.04	1.7	3.073	1.749	4.19	7.83	0.85	14.6	0.8221	Normal
4	3.04	1.7	2.936	1.645	3.77	7.45	0.85	14.72	0.8283	Normal
5	2.81	2.1	2.771	2.030	4.41	7.62	0.95	13.16	0.6806	Normal
6	4.14	1.17	4.202	1.134	3.72	9.66	0.5	25	0.9692	Normal
7	4.1	3.32	4.0155	3.2479	10.17	11.45	0.97	12.89	0.7729	Normal
8	2.9	1.82	2.8902	1.6936	3.83	7.43	0.87	14.41	0.8343	Normal
9	2.38	1.9	2.4184	1.7017	3.21	6.55	0.94	13.36	0.7819	Normal
10	6.24	4.27	6.28	3.97	18.43	20.06	0.57	21.83	0.7	Abnormal

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